



CHALMERS
UNIVERSITY OF TECHNOLOGY

Exposure-response modeling improves selection of radiation and radiosensitizer combinations

Downloaded from: <https://research.chalmers.se>, 2026-04-03 07:54 UTC

Citation for the original published paper (version of record):

Cardilin, T., Almquist, J., Jirstrand, M. et al (2022). Exposure-response modeling improves selection of radiation and radiosensitizer combinations. *Journal of Pharmacokinetics and Pharmacodynamics*, 49(2): 167-178. <http://dx.doi.org/10.1007/s10928-021-09784-7>

N.B. When citing this work, cite the original published paper.



Exposure-response modeling improves selection of radiation and radiosensitizer combinations

Tim Cardilin^{1,2} · Joachim Almquist^{1,6} · Mats Jirstrand¹ · Astrid Zimmermann³ · Floriane Lignet⁴ · Samer El Bawab⁴ · Johan Gabrielsson⁵

Received: 30 March 2021 / Accepted: 19 September 2021
© The Author(s) 2021

Abstract

A central question in drug discovery is how to select drug candidates from a large number of available compounds. This analysis presents a model-based approach for comparing and ranking combinations of radiation and radiosensitizers. The approach is quantitative and based on the previously-derived Tumor Static Exposure (TSE) concept. Combinations of radiation and radiosensitizers are evaluated based on their ability to induce tumor regression relative to toxicity and other potential costs. The approach is presented in the form of a case study where the objective is to find the most promising candidate out of three radiosensitizing agents. Data from a xenograft study is described using a nonlinear mixed-effects modeling approach and a previously-published tumor model for radiation and radiosensitizing agents. First, the most promising candidate is chosen under the assumption that all compounds are equally toxic. The impact of toxicity in compound selection is then illustrated by assuming that one compound is more toxic than the others, leading to a different choice of candidate.

Keywords Radiosensitizer · Tumor Static Exposure · Treatment optimization · Tumor growth model · Drug selection

Introduction

Radiotherapy is a cornerstone of modern oncology, and is frequently given in conjunction with chemical treatments to improve efficacy [1, 2]. Radiosensitizers are a class of chemical agents designed to enhance the radiation effect, e.g. by interfering with the cell's repair of radiation-

induced DNA damage [3]. During preclinical development of novel compounds, including radiosensitizers, a central question is how to select the most promising compounds from a large number of candidates [4–6]. Proper assessment of radiation and radiosensitizer combinations requires studies of efficacy as well as toxicology and adverse effects [7]. All compounds and doses cannot be tested in vivo—for reasons of time, resources, and ethics [8]. Experimental studies must therefore be supported by cheaper alternatives such as computer modeling and simulations [9, 10].

Numerous quantitative models have been developed to describe the effects of radiotherapy on tumors, with or without chemical intervention [11–14]. These models range from the simple, yet ubiquitous, linear-quadratic model of radiobiology [15], to complex systems pharmacology models that include particular pathways and processes (such as the cell cycle) that are relevant to the given treatment [16, 17]. In radiation oncology, models of Tumor Control Probability (TCP)—defined by whether a given radiation dose controls or eradicates an irradiated tumor—are commonly employed alongside Normal Tissue Complication Probability (NTCP) models that quantify toxicology and adverse risks [18–20].

✉ Tim Cardilin
tim.cardilin@fcc.chalmers.se

¹ Fraunhofer-Chalmers Centre, Chalmers Science Park, Gothenburg, Sweden

² Department of Mathematical Sciences, Chalmers University of Technology and University of Gothenburg, Gothenburg, Sweden

³ Translation Innovation Platform Oncology, Merck KGaA, Darmstadt, Germany

⁴ Translational Medicine, Merck KGaA, Darmstadt, Germany

⁵ Meddoor AB, Gothenburg, Sweden

⁶ Present Address: Clinical Pharmacology and Quantitative Pharmacology, Clinical Pharmacology & Safety Sciences, BioPharmaceuticals R&D, AstraZeneca, Gothenburg, Sweden

Models have also been developed to describe the effects of radiotherapy on tumor volume over time. Watanabe et al. [21] proposed a radiation model with gradual cell death in response to single-dose treatment, and used it to describe tumor growth over time in rat rhabdomyosarcoma and in patients with metastatic brain tumors. More recently, Husband et al. developed and evaluated radiation models that describe tumor growth and survival in patient-derived xenograft mice for diffuse intrinsic pontine glioma [22].

In two earlier papers, we developed models that describe tumor growth in xenograft mice receiving radiotherapy and neoadjuvant radiosensitizing treatment [23, 24]. We also introduced the Tumor Static Exposure (TSE) concept—a model-based prediction of all combinations of radiation doses and radiosensitizer concentrations that result in tumor regression. However, these models only consider a single radiosensitizing compound and can therefore not fully illustrate the utility of the TSE concept in aiding the drug selection process.

In this paper, we use TSE to compare and rank three different combinations of radiation and radiosensitizing agents. One of our earlier models is used with data from a xenograft study involving radiotherapy administered alone or together with either of the radiosensitizers. The compounds are ranked by weighing efficacy (measured using TSE) against toxicity. Two different toxicological models are considered: a simple, linear model; and a more complex NTCP model adjusted to account for radiosensitizing treatment [25]. We also introduce the concept of Tumor Shrinkage Exposures, which can be used if tumor stasis is insufficient and tumor shrinkage with a particular rate is desired.

Methods

Experimental data are first described. Then, a previously-developed tumor model used to describe radiation and radiosensitizer combination therapies is summarized. Thereafter, a method for comparing and ranking radiation and radiosensitizer combinations, based on TSE, is presented. The method optimizes a given cost function, used to describe, *e.g.*, toxicity and other adverse effects, along the TSE curve. Finally, computational aspects of the non-linear mixed-effects modeling approach are provided.

Experimental data

Pharmacodynamic data were generated in FaDu xenograft mouse models treated with radiation either alone or together with one of three early-discovery radiosensitizing compounds, henceforth referred to as compounds A₁, A₂, and A₃. A total of 54 female mice were divided into six

treatment groups with nine mice in each group: (A) vehicle control, (B) monotherapy with radiation (2 Gy per dose), (C) combination therapy with radiation (2 Gy per dose) and compound A₁ (100 mg/kg per dose), (D) combination therapy with radiation (2 Gy per dose) and compound A₂ (25 mg/kg per dose), (E) combination therapy with radiation (2 Gy per dose) and compound A₂ (100 mg/kg per dose), and (F) combination therapy with radiation (2 Gy per dose) and compound A₃ (20 mg/kg per dose). Doses were given once per day Mon–Fri for 6 weeks.

Exposure data were generated in FaDu xenograft models for the compounds A₁, A₂, and A₃. Single doses of the compounds A₁, A₂, and A₃ were given orally to 16 animals divided into four treatment groups with four mice in each group: compound A₁ (100 mg/kg), compound A₂ (25 mg/kg), compound A₂ (100 mg/kg) and compound A₃ (20 mg/kg). Drug concentration in plasma was measured after 1, 2, and 6 h.

Experiments were approved in accordance with German animal welfare regulations by the Regierungspräsidium Darmstadt, Hessen, Germany (protocol registration numbers DA 4/Anz. 397 and DA 4/Anz. 398).

Tumor model for radiation and radiosensitizer combination treatment

We use a previously-developed radiation model (Fig. 1) to describe tumor growth following treatment with radiation and radiosensitizing agents [23]. The model consists of a main compartment V_1 representing proliferating cancer cells, three damage compartments V_2 , V_3 , and V_4 , that all

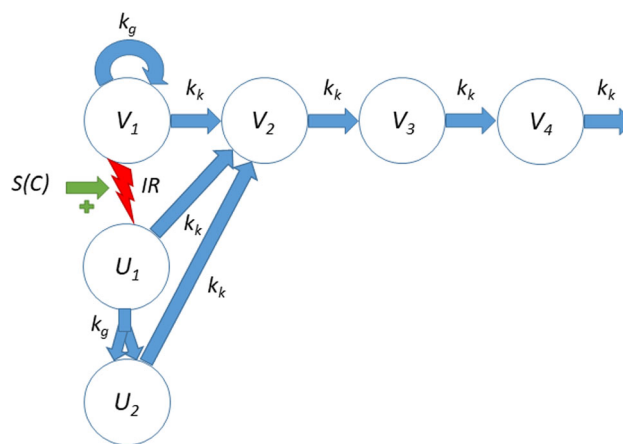


Fig. 1 Tumor model used to describe combination therapy with ionizing radiation (IR) and radiosensitizer compounds. Cancer cells in compartment V_1 proliferate with rate k_g and are eliminated with rate k_k . Dying cells are transferred through three damage compartments V_2 , V_3 and V_4 . Lethally irradiated cells are moved to a radiation-damage compartment U_1 where they are allowed up to one more cell division, before dying. The compartment U_2 represents irradiated cells after one cell division that can no longer divide

dying cells traverse, and two radiation compartments U_1 , and U_2 , that allow irradiated cells up to one more cell division before dying. Irradiated cells are instantaneously transferred from V_1 to U_1 . The fraction of proliferating cells that is transferred is based on the well-established linear-quadratic formula from radiobiology [14, 15]. Moreover, the presence of a radiosensitizing agent is accounted for via an increase in the number of lethally irradiated cells depending on the plasma concentration of the radiosensitizer at the time of irradiation. A high plasma concentration leads to a greater transfer of cells from V_1 to U_1 . The model also incorporates natural cell death, meaning that some cells traverse the damage compartments even for untreated tumors.

The tumor model is described by the following differential equations

$$\begin{aligned} \frac{dV_1}{dt} &= k_g V_1 - k_k V_1 \\ \frac{dV_2}{dt} &= k_k V_1 + k_k U_1 + k_k U_2 - k_k V_2 \\ \frac{dV_3}{dt} &= k_k V_2 - k_k V_3 \\ \frac{dV_4}{dt} &= k_k V_3 - k_k V_4 \\ \frac{dU_1}{dt} &= -k_g U_1 - k_k U_1 \\ \frac{dU_2}{dt} &= 2k_g U_1 - k_k U_2 \end{aligned} \tag{1}$$

where k_g is the growth rate of proliferating cancer cells, and k_k the kill rate of cancer cells which is assumed to be the same for all compartments. The use of growth rate k_g and the presence of the factor two in the transfer from U_1 to U_2 describes that cell division occurs between these states and therefore twice as many cells enter U_2 than leave U_1 .

Radiation treatment is implemented as sudden transfer between compartments V_1 and U_1 , corresponding to an instantaneous transfer of cells with the fraction given by $(1 - SF(D_{IR}, C_j))$. Here, $SF(D_{IR}, C_j)$ is the surviving fraction of proliferating cancer cells given a radiation dose D_{IR} and concurrent drug plasma concentration C_j of compound A_j . Mathematically this can be described by the two equations

$$\begin{aligned} V_1(t_i^+) &= V_1(t_i^-) - (1 - SF(D_{IR}(t_i), C_j(t_i))) V_1(t_i^-), \\ U_1(t_i^+) &= U_1(t_i^-) + (1 - SF(D_{IR}(t_i), C_j(t_i))) V_1(t_i^-), \end{aligned} \tag{2}$$

where t_i denotes the times of irradiation, and t_i^- and t_i^+ can be interpreted as times just before and after irradiation. Note that radiation dose is given in terms of Gray (Gy), which is absorbed dose measured in joules per kilogram, *i.e.*, the radiation dose is normalized with respect to animal weight and hence plays the role of exposure to radiation. The surviving fraction is given by

$$SF(D_{IR}, C_j) = \exp[-(1 + a_j C_j)(\alpha D_{IR} + \beta D_{IR}^2)] \tag{3}$$

where α and β are the linear and quadratic coefficients associated with radiation DNA damage, and a_j is the pharmacodynamic parameter associated with the radiosensitizing capabilities of compound A_j . The initial conditions for the system are given by

$$V_i(0) = V^0 \left(\frac{k_k}{k_g}\right)^{i-1}, \quad U_i(0) = 0, \tag{4}$$

where V^0 is the initial volume of the main compartment. With these initial conditions, untreated tumors grow exponentially with net growth rate $k_g - k_k$ [26]. The total tumor volume, V_{total} , is obtained as the sum of all compartments

$$V_{total} = V_1 + V_2 + V_3 + V_4 + U_1 + U_2 \tag{5}$$

Comparing combinations of radiation and radiosensitizers

In the case study, the goal is to select one of three radiosensitizing agents for further experimental study. We propose a model-based approach that evaluates combinations based on how easily tumor regression is achieved, relative to toxicological or other adverse effects. The model described in the previous section is calibrated to data and then used to derive TSE curves for each radiosensitizing agent. Cost functions are introduced to describe toxicology and other potential costs associated with treatment, and an optimization problem is formulated to minimize the cost subject to the constraint that the tumor does not grow, *i.e.*, that the exposure is on or above the TSE curve.

The Tumor Static Concentration (TSC) and TSE concepts have been introduced and used in several earlier papers [26–29]. The TSC curve corresponding to a particular combination therapy consists of all pairs (C_1, C_2) of plasma concentrations for which a maintained exposure leads to tumor stasis. In particular, maintaining exposure levels above the TSC curve leads to tumor regression. The TSE concept is a generalization of TSC that allows for treatments for which concentrations are unknown or not applicable, such as radiotherapy. The TSE curve for the model given in Eqs. 1 and 2 has previously been derived (see [23]). The curve consists of combinations of daily radiation doses and average radiosensitizer concentrations such that the tumor is kept in approximate stasis.

In the Results section, the calibrated tumor model is used to generate TSE curves for combination therapy with radiation and each of three radiosensitizers, which we denote A_1 , A_2 , and A_3 . We propose the following

procedure for comparing and ranking combinations, while accounting for toxicity and other adverse effects.

For each treatment combination, introduce an associated cost function $\Psi(E_1, E_2)$, where E_1 and E_2 refer to general exposure metrics. In our case study $E_1 = D_{IR}$ is radiation dose, and $E_2 = C_j$ is the concurrent plasma concentration of radiosensitizer A_i . Alternatively, nondimensional exposures could be defined by $E_1 = D_{IR}/D_{ref}$ and $E_2 = C_j/C_{ref,j}$, where D_{ref} and $C_{ref,j}$ are reference exposures. The cost function is a way to measure the toxicity of the combination, although other kinds of costs could also be included. Ψ is an increasing function, reflecting that a larger value corresponds to higher cost/toxicity.

In the simplest case, Ψ is linear function and is given by

$$\Psi(E_1, E_2) = pE_1 + qE_2, \quad (6)$$

where p/q is the relative toxicity of the two compounds, assumed to be constant. Equation 6 assumes that toxicity increases linearly with exposure and is additive. Exposure pairs of equal costs, *i.e.*, the level curves $\Psi(E_1, E_2) = \text{constant}$, are in this case lines with slope $-p/q$, with E_1 and E_2 are on the horizontal and vertical axes, respectively. An example of a TSE curve and a level set of the cost function is shown in Fig. 2.

As an example of a more intricate cost function, we utilize the established framework around NTCP [18, 19]. Such models are commonly used to describe the probability of adverse events following radiation treatment [30–32]. A widely used model for NTCP is the Lyman–Kutcher–Burman (LKB) model which defines the NTCP as

$$NTCP(t) = \frac{1}{\sqrt{2\pi}} \int_{-\infty}^t e^{-x^2/2} dx, \quad (7)$$

where the variable t is defined by

$$t = \frac{D_{eff} - TD_{50}}{mTD_{50}}, \quad (8)$$

and where D_{eff} is the effective dose, which accounts for non-uniform dose distribution, TD_{50} is the dose associated with 50% complication risk, and m is a slope parameter for the sigmoidal curve [25, 33]. From these equations we can see that a larger value of D_{eff} corresponds to a larger value for t , which in turns means a greater risk of complications.

The key question when defining a cost function for radiation and radiosensitizer combinations is how to introduce radiosensitizing treatment into the NTCP model. Since TD_{50} is a typical measure of radiation sensitivity, we propose to let the radiosensitizer modulate this parameter and thereby increase the risk of complications. Assuming an exponential sigmoidal modulation function gives a new definition of t ,

$$t = \frac{D_{eff} - TD_{50}I(C)}{mTD_{50}I(C)} \quad (9)$$

where $I(C)$ is an exponential inhibitory function with parameter k_s [34]:

$$I(C) = \exp(-k_s C). \quad (10)$$

We can thus use the NTCP model as a cost function with exposures $E_1 = D_{eff} = D$ (total radiation dose) and $E_2 = C$, where C is the radiosensitizer concentration at the time of irradiation. Note that NTCP depends on the exposures E_1 and E_2 only through the variable t . Therefore, exposure combinations with equal complication risk have the same value for t . Thus, solving for D_{eff} in Eq. 9 gives the expression for equal cost

$$D_{eff} = (1 + mt)TD_{50} \exp(-k_s C). \quad (11)$$

Equation 10 describes a sigmoidal relationship between exposure pairs (D_{eff}, C) with equal cost.

Equipped with a cost function, we search along the TSE curve for the exposure pair with the lowest cost. Repeating this procedure for each combination gives a sequence of lowest costs, each corresponding to a different combination therapy. These values can then be used to compare and rank combination therapies.

The procedure for comparing and ranking combinations is summarized below:

- (1) Choose a suitable tumor model given available data and calibrate the model to obtain parameter estimates
- (2) Compute the TSE curves and insert the estimated parameter values

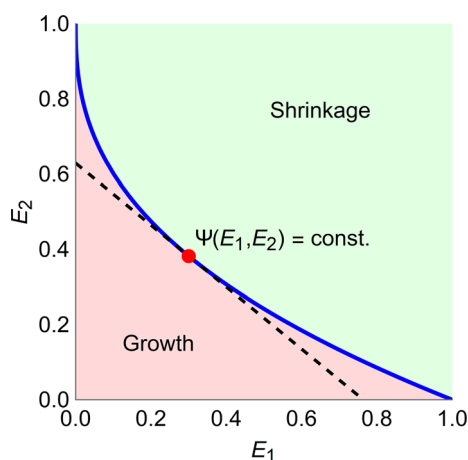


Fig. 2 TSE curve for two compounds with exposures E_1 and E_2 (blue). Exposure pairs above the curve give rise to tumor shrinkage, whereas exposure pairs below the curve result in tumor growth. A level set where the cost Ψ is constant is shown in black, dashed, with the corresponding Ψ^* (color figure online)

- (3) Choose an appropriate cost function Ψ for each combination and find the minimum cost Ψ^* along the TSE curve
- (4) Repeat Steps 1–3 for each drug combination
- (5) Compare Ψ^* across combination therapies and choose the combination with the lowest cost

Tumor shrinkage exposures

TSE curves are based on the requirement of tumor stasis. This is valuable since the curve divides the exposure plane into regions of tumor growth and tumor shrinkage. However, in practice, tumor shrinkage may not be enough and one can therefore look at tumor shrinkage with different rates. This leads to a generalization of TSE called tumor *shrinkage* exposure (TSE_ρ) where ρ is the relative change in volume for a given time unit, $\rho = \frac{V(t_1) - V(t_2)}{V(t_1)}$, where $t_2 > t_1$ are two time point, and $t_2 - t_1$ is the chosen time unit. In particular, TSE_0 is the regular TSE curve, and $TSE_{0.5}$ requires that the tumor shrinks by 50% of its size every for every unit of time that elapses. TSE_ρ is derived analogously to TSE, with the difference that the growth rate is set to a constant different from zero. The concept of TSE_ρ curves is illustrated for the case study in the [Results](#) section.

Computational methods

The tumor model was calibrated to xenograft data using a nonlinear mixed-effects approach based on the first-order conditional estimation (FOCE) method in a computational framework developed at the Fraunhofer-Chalmers Research Centre for Industrial Mathematics (Gothenburg, Sweden) and implemented in Mathematica (Wolfram Research) [35]. Exposure data for compounds A_1 , A_2 , and A_3 were described using one-compartment pharmacokinetic models. Model evaluation was based on goodness-of-fit, empirical Bayes estimates, and residual analysis.

Lognormal distributed between-subject variability was added to the initial volume of the main compartment V_0 and the growth rate k_g . Residual errors were assumed to be proportional to tumor volume with zero mean and variance σ_v^2 . As done previously, the ratio between α and β was set to 10 [23, 36].

Results

First, the results of fitting the tumor model to the experimental data are presented. Then, TSE curves corresponding to combination therapy with radiation and each of the three radiosensitizers A_1 , A_2 , and A_3 , are computed. Finally, the

procedure for comparing and ranking combinations is illustrated for two toxicological settings.

Tumor model for radiation and radiosensitizer combination treatment

Exposure profiles for each of the three radiosensitizers A_1 , A_2 , and A_3 were described by standard one-compartment pharmacokinetic models, with parameter estimates given in Table 1. Simulated PK profiles used to drive the pharmacodynamic tumor model are shown in Fig. 3. The exposure of compound A_1 (green) was approximately ten times lower than the exposures of compounds A_2 (blue) and A_3 (purple).

The tumor model adequately described observed data from each of the six treatment groups. Examples of individual fits for each treatment group are shown in Fig. 4. In the vehicle group, tumor growth was approximately exponential during the observed time period. Tumors treated with radiation monotherapy reached approximate stasis during treatment and in some cases showed signs of regression. Tumors treated with radiation and compound A_1 combination therapy exhibited significant regression and in most cases the tumors were eradicated. Tumors treated with radiation and compound A_2 showed significant regression with the lower dose (25 mg/kg) and in most cases tumor eradication with the higher dose (100 mg/kg). Lastly, tumors treated with radiation and compound A_3 also exhibited tumor eradication in most instances. Visual predictive checks for the tumor model can be found in Supplemental Information S1.

The parameter estimates from fitting the tumor model simultaneously to all treatment groups are given in Table 2. The net growth rate $k_g - k_k = 0.16/\text{day}$ corresponds to an average doubling time of 4.3 days for the vehicle group. System and radiation parameters were estimated with good precision, whereas drug parameters were estimated with lower but still acceptable precision (RSE < 40%).

TSE curves for radiation and radiosensitizer combinations

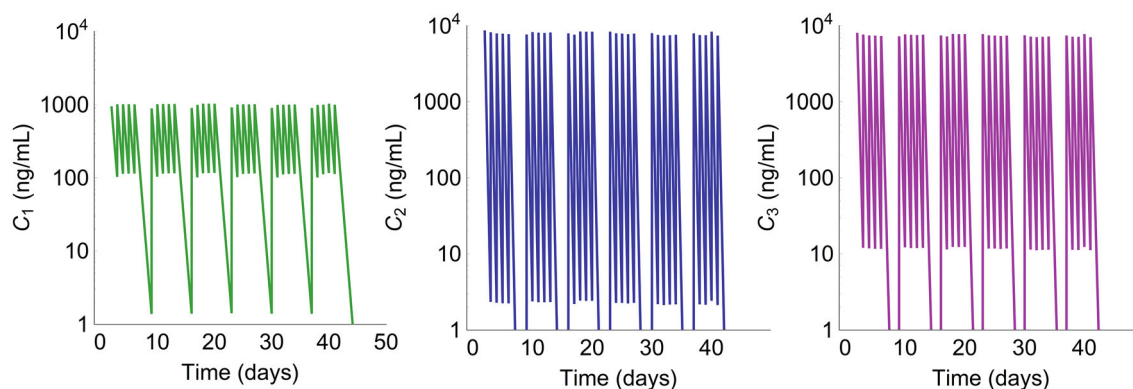
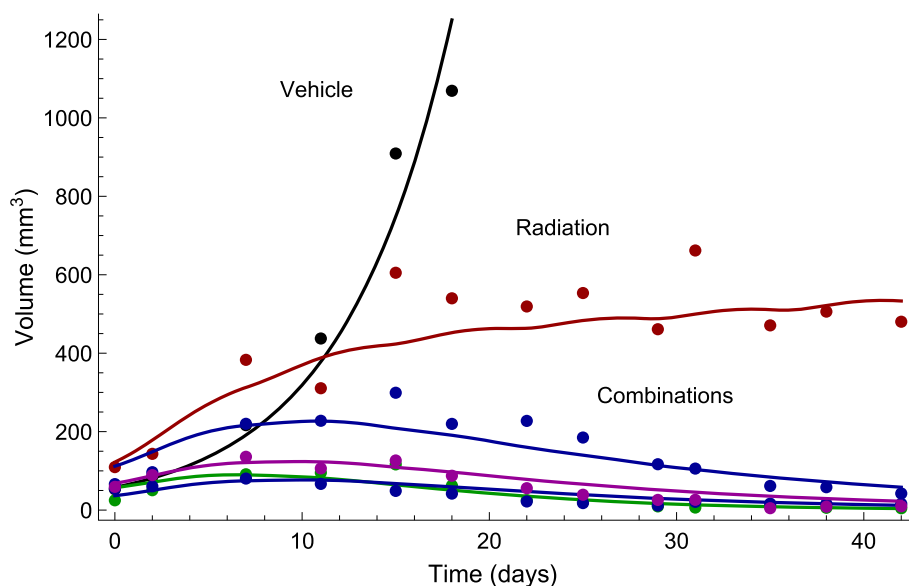
Following the same principles as in [23] the following expression for the TSE_ρ curves was derived

$$D_{IR} = \frac{-(\alpha + a_j \alpha C_j) + \sqrt{(\alpha + a_j \alpha C_j)^2 + 4(\beta + a_j \beta C_j)(k_g T - k_k T + \log(1 - \rho))}}{2(\beta + a_j \beta C_j)} \quad (12)$$

where D_{IR} is the radiation dose given every T days, and C_j is the plasma concentration of A_j at the instance of irradiation. The details can be found in Supplemental Information S2.

Table 1 Parameter estimates for the one-compartment pharmacokinetic models describing exposure to the compounds A₁, A₂, and A₃ in terms of plasma concentration

Parameter	Compound	Population median (RSE%)	Between-subject variability ^a (RSE%)
k_e (/h)	A ₁	0.092 (9)	63 (15)
	A ₂	0.35 (7)	18(13)
	A ₃	0.27 (9)	6 (13)
V (L/kg)	A ₁	110 (8)	26 (14)
	A ₂	12 (8)	14 (17)
	A ₃	2.6 (6)	2 (21)
σ_e^b (%)	A ₁	14 (25)	–
	A ₂	15 (22)	–
	A ₃	27 (20)	–

^aCalculated as $\sqrt{\omega_{ii}^2} \times 100$ ^bIntra-individual variability**Fig. 3** Simulated PK profiles for compounds A₁, A₂, and A₃ with corresponding plasma concentrations C₁, C₂, and C₃. Doses of 100 mg/kg (compounds A₁ and A₂) or 20 mg/kg (compound A₃) were given 5 days a week for 6 weeks**Fig. 4** Examples of individual fits for each of the six treatment groups: vehicle (black), radiation monotherapy with 2 Gy per dose (red), combination therapy with radiation and A₁ at 100 mg/kg per dose (green), combination therapy with radiation and A₂ at 25 mg/kg or 100 mg/kg per dose (blue), and combination therapy with radiation and A₃ at 20 mg/kg per dose (purple) (color figure online)

The TSE curves for the three combination therapies involving radiation and one of the radiosensitizing agents A₁, A₂, and A₃, were computed by inserting the parameter

estimates from Table 2 into Eq. 12, using $T = 1$ day to indicate daily dosing. The resulting TSE curves are shown in Fig. 5.

Table 2 Parameter estimates for the tumor model describing the effects of radiation and radiosensitizer combination therapy

Parameter	Population median (RSE%)	Between-subject variability ^a (RSE%)	Description
k_g (/day)	0.50 (5)	53 (2)	Natural growth rate
k_k (/day)	0.34 (5)	–	Natural kill rate
V^0 (mm ³)	26.0 (8)	7 (11)	Initial volume of main compartment
α (/Gy)	0.11 (6)	–	Linear radiation parameter
β (/Gy ²)	0.011 (6)	–	Quadratic radiation parameter
a_1 (mL/ μ g)	0.27 (33)	–	Pharmacodynamic parameter of A_1
a_2 (mL/ μ g)	0.038 (36)	–	Pharmacodynamic parameter of A_2
a_3 (mL/ μ g)	0.028 (35)	–	Pharmacodynamic parameter of A_3
σ_V^b (%)	28.0 (3)	–	Proportional standard error

^aCalculated as $\sqrt{\omega_{ii}^2} \times 100$

^bIntra-individual variability

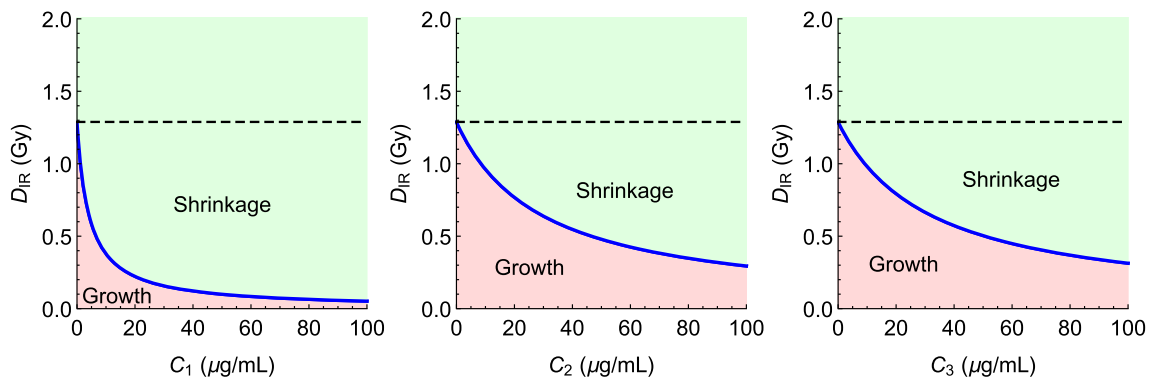


Fig. 5 TSE curves for combinations of radiation and radiosensitizers A_1 (left), A_2 (middle) and A_3 (right) obtained by inserting the parameter estimates from Table 2 into Eq. 12. TSE curves are shown in blue. Regions above and below the curves correspond to

combination pairs that result in tumor shrinkage, or tumor growth, respectively. The dashed reference lines indicate the daily radiation dose required for tumor shrinkage during monotherapy

The TSE value for radiation monotherapy was estimated to 1.3 Gy, meaning that for the median individual a daily dose of 1.3 Gy would be sufficient for approximate tumor stasis. Since the compounds A_1 , A_2 , and A_3 , have no monotherapy effect, they have no TSE values. Instead, the TSE curves asymptotically approach the concentration axis as the plasma concentrations of the compounds approach infinity. The TSE curve for combinations of radiation and compound A_1 (left) exhibits the largest curvature. Indeed, that TSE curve associated with A_1 lies strictly below the TSE curve for the other two combination therapies.

Comparing combinations of radiation and radiosensitizers

The procedure outlined in the Methods section is applied to compare and rank the three combination therapies for two toxicological models. Using the first model, we consider two scenarios: one based on the assumption that all

radiosensitizers are equally toxic, and one where the toxicity of compound A_1 is increased tenfold. The cost functions are given by

$$\Psi(D_{IR}, C_j) = pD_{IR} + q_j C_j \quad (13)$$

where D_{IR} is the radiation dose with toxicity coefficient p , and C_j is the plasma concentrations of compound A_j with toxicity coefficient q_j . First, assuming that all test compounds are equally toxic means that $q_1 = q_2 = q_3$. The costs associated with each combination pair (D_{IR}, C_j) on the corresponding TSE curve are illustrated in Fig. 6 (left). The parameter s indicates the location along the TSE curve with $s = 0$ corresponding to radiation monotherapy, and $s = 1$ corresponding to monotherapy with the radiosensitizer. Note that, for the particular tumor model in this case study, the radiosensitizers have no monotherapy effect, which means that $s = 1$ corresponds to an infinitely large exposure of the radiosensitizer and therefore also an infinite cost/toxicity. The parametrization has been performed

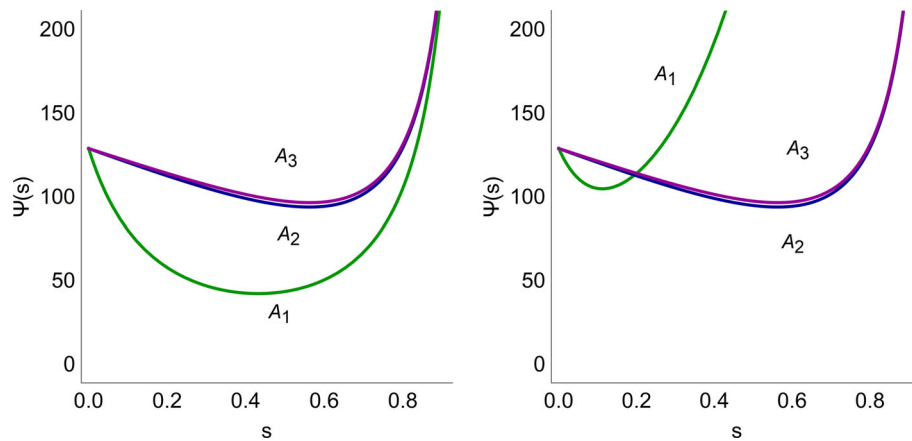


Fig. 6 Hypothetical costs Ψ for different combinations along the TSE curves in Fig. 5 for combination therapy with radiation and radiosensitizers A_1 (green), A_2 (blue), and A_3 (purple). The left plot assumes that all three compounds are equally toxic, whereas in the

right plot the toxicity of A_1 (green) has been increased by a factor ten. The parameter s represents the position on the TSE curve with $s = 0$ corresponding to radiation monotherapy and $s = 1$ monotherapy with compound A_j (color figure online)

such that $s = 0.5$ corresponds to $C_j = 25 \mu\text{g/mL}$. This is an arbitrary scaling of the parametrization that does not affect the optimization problem and is performed only to make the figures easier to interpret. This amounts to the parametrization given in Eq. 14 below

$$\Psi(s) = pD_{IR} \left(\frac{25s}{1-s} \right) + q_j \frac{25s}{1-s} \quad (14)$$

with $D_{IR}(C_j)$ given as in Eq. 12.

The cost coefficients were set to $p = 100/\text{Gy}$ and $q_j = 1 \text{ mL}/\mu\text{g}$. Figure 6 (left) shows that combination therapy with A_1 has the lowest cost (for $s \approx 0.5$). We then consider the second scenario, where the toxicity of A_1 has been increased by a factor ten, $q_1 = 10 \text{ mL}/\mu\text{g}$, which is illustrated in Fig. 6 (right). A_1 is no longer the best treatment option, since A_2 and A_3 both have lower costs (occurring at $s \approx 0.6$).

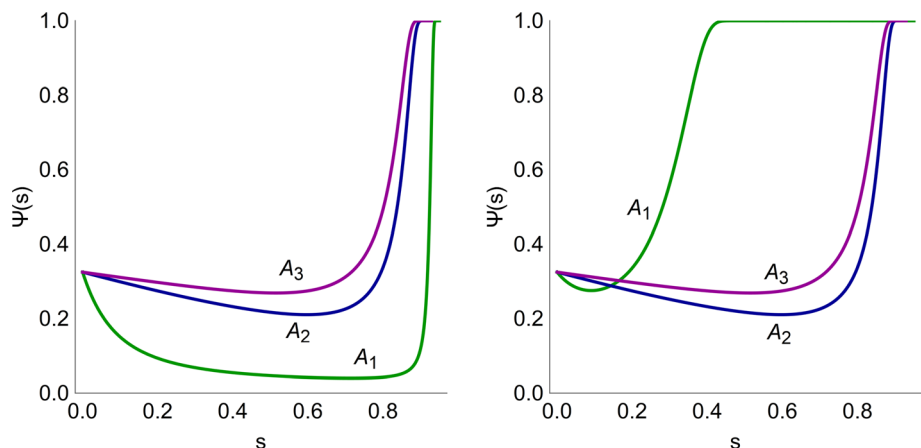
Figure 7 shows the results using the more complex NTCP model as cost function. For this model, we use values of $TD_{50} = 50 \text{ Gy}$ and $m = 0.5$ to describe radiation

treatment and set the radiation parameter k_s to $0.02 \text{ mL}/\mu\text{g/day}$ for all three radiosensitizers. Similar to the case with a linear cost function, A_1 , which is the most efficacious, has the lowest cost. Compared with the linear case, the value of the radiosensitizer parameter k_s for A_1 would need to be decreased approximately tenfold for another radiosensitizer to become the most promising candidate. The parametrization of the cost function along the TSE curve, with parameter s going from $s = 0$ (radiotherapy) to $s = 1$ (radiosensitizer monotherapy) is given in Eq. 15.

$$\Psi(s) = \text{NTCP} \left(\frac{D_{IR} \left(\frac{25s}{1-s} \right) - TD_{50} I \left(\frac{25s}{1-s} \right)}{TD_{50} I \left(\frac{25s}{1-s} \right)} \right) \quad (15)$$

where D_{IR} is given by Eq. 12, I is given by Eq. 10, and NTCP is given by Eq. 9.

Fig. 7 Hypothetical costs Ψ using the NTCP model (Eq. 15) for radiation and radiosensitizer combinations, A_1 (green), A_2 (blue), and A_3 (purple), along the TSE curves in Fig. 5. The left plot assumes equal toxicity, whereas the right plot assumes a tenfold increase for radiosensitizer A_1 (color figure online)



Tumor shrinkage exposures

Figures 5 and 6 are used to minimize the cost of combination therapy with the respect to the TSE curve, *i.e.*, while making sure the tumor is not growing. As pointed out in the Methods section, it is also possible to require that the tumors shrink at a specified rate, by introducing the TSE_ρ curves. TSE_ρ curves are illustrated in Fig. 8 for combinations of radiation and compound A_2 , assuming a linear cost function as in Fig. 6. The three TSE_ρ curves (blue) consists of exposure pairs (D_{IR}, C_j) that keep the tumor in stasis, shrink the tumor to half its size, and shrink the tumor to one eighth of its size, respectively.

For each TSE_ρ curve, it is possible to minimize the costs of combination therapy following the same procedure as described earlier. Thus, for a given combination, consider how the minimum cost, Ψ_{\min} , varies depending on how quickly the tumor is required to shrink, *i.e.*, ρ . Figure 9 depicts this scenario for combinations of radiation and the three radiosensitizers as a function of the parameter $1/1 - \rho$ under the assumption that all compounds are equally toxic. Note that combination therapy with A_1 (green) always has the lowest cost.

Discussion

Recent decades have seen a growing focus on combination therapies as a way to combat resistances and to obtain synergistic effects [37, 38]. Our analysis of radiotherapy and radiosensitizer combinations in this paper is focused on the latter, while also addressing toxicity and side-effects. As with any model-based analysis, good predictions and

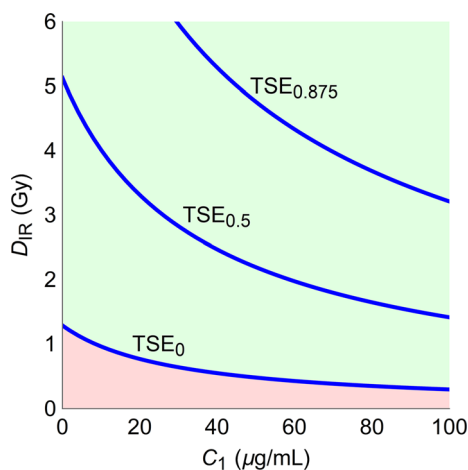


Fig. 8 Examples of TSE curves for combinations of radiation and compound A_2 . TSE_1 , TSE_2 , and TSE_8 corresponding to shrinkage rates that keep the tumor in stasis, reduce the tumor to half its size, and reduce the number of proliferating tumor cells to one eighth of its size with each daily dose, respectively

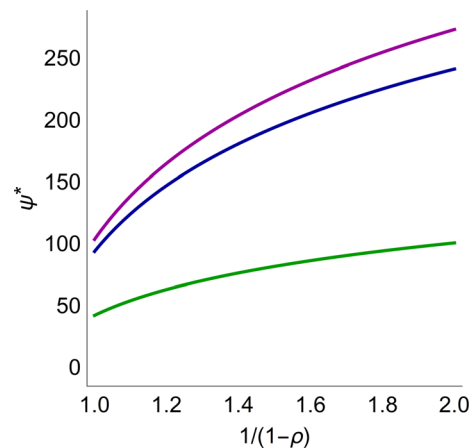


Fig. 9 Minimal costs ψ^* as a function of relative shrinkage rate ρ for combinations of radiation and A_1 (green), A_2 (blue), and A_3 (purple). Here

$$\frac{1}{1 - \rho} = V_1(t_1)/V_1(t_2)$$

is a more natural parameter such that the minimal cost is an increasing function of the parameter. The figure shows that, for any value of ρ , the compound A_1 has the lowest cost, since the curves never intersect

results are contingent not only on data [39], but also on sound modeling methodology [40, 41]. Details on this topic, particularly in the context of oncology, can be found *e.g.* in several papers by Mould et al. [42–44]. The remainder of this discussion considers, in order: the mathematical tumor model, the resulting TSE curves and concepts, and, finally, our proposed optimization and ranking procedure for radiation and radiosensitizer combinations.

Tumor model for radiation and radiosensitizer combinations

The model used in this paper is based on an earlier model (see [23]), with two minor differences. As in [24] an exponential growth function was favored over logistic growth, since it proved sufficient to describe vehicle data and no plateaus in tumor volumes were observed. Secondly, we assumed no monotherapy effect for the radiosensitizers, which is expected to be negligible given that the compounds interact with the repair mechanisms of DNA damage induced by irradiation.

The growth and kill rates were estimated to similar values to those in [23, 24], and the net growth rate $k_g - k_k$ of 0.15/day, corresponds to a doubling time of 4–5 days, which is similar to other models [21, 23, 26, 27, 45]. The estimated α and β values of 0.11/Gy and 0.0011/Gy² are in line with reported ranges of 0.02 – 0.2 for α and 0.001 – 0.6 for β [46]. Model parameters were estimated

with reasonable precision, although the radiosensitizer parameters a_i had somewhat lower precision (RSE% \approx 35), which is partially explained by the fact that each a_i is only informed by one or two treatment groups, whereas other model parameters are informed by all data.

Like many tumor models used preclinically, our model contains a sequence of damage compartments [26, 27, 29, 45], and can be viewed as a combination of these models with the linear-quadratic model in radiobiology [15], or compartment radiation models that implement the linear-quadratic model with delay [21, 22]. Compared with systems pharmacology models for radiation and chemical combinations, *e.g.*, Checkley et al. [17], Kosinsky et al. [16], our model is simpler and, although less mechanistic, can be calibrated to standard xenograft data.

TSE curves for radiation and radiosensitizer combinations

TSC and TSE have been developed and applied in a series of papers [23, 26, 27, 29]. They are tailored specifically to cancer treatments (single-agents or combinations), and are connected to qualitative behavior (tumor growth or shrinkage) of the disease as well as synergy, unlike general models such as the isobologram [47, 48] and the half-maximal effect curve [49] which focus only on synergy. In radiation oncology, TCP models are used to assess probabilities of tumor eradication, recurrence, or emergence of metastases [20, 50], which is similar to TSE in that it also aims to control or destroy cancer cells.

In our analysis, greatest synergy occurred with radiosensitizer A_1 , which can also be seen from the curvatures of the TSE curves (Fig. 5). This happens because although observed tumor growth was similar across combinations, exposure levels were approximately ten times lower for compounds A_1 (see Fig. 3). However, proper assessment requires consideration not only of efficacy, but joint consideration of efficacy and toxicity.

Comparing combinations of radiation and radiosensitizers

In our case study involving radiation and three radiosensitizing agents, Fig. 6 (left) shows that radiosensitizer A_1 is the superior radiosensitizer given that all compounds are equally toxic, which holds for either cost function. Moreover, Fig. 6 (right) shows that the toxicity of A_1 would have to be increased tenfold over A_2 and A_3 for another radiosensitizer to become preferable. This result held true for both cost functions. However, since the NTCP model is nonlinear and contains multiple sigmoidal functions, these results depend on the chosen parameter values.

Our proposed method evaluates combinations of radiation and radiosensitizers by the ability to induce tumor regression relative to toxicity. Two toxicity functions, or cost functions are considered: one linear, and one based on NTCP. The former approach was also considered in [29] to find an optimal combination for two anticancer compounds. A similar analysis can be found in [51] where phase one clinical data were used to construct a toxicity function with linear terms as well as a quadratic term to penalize combination treatment.

In radiotherapy, TCP and NTCP models are often combined to optimize treatment [52, 53]. In our analysis, the tumor model together with TSE appear instead of a TCP model, which we consider in conjunction with the commonly used Lyman NTCP model [25]. Here, we note similar results using a linear model and an NTCP model (see Figs. 6, 7) although the sigmoidal nature of the NTCP model produced somewhat flatter cost around the minima, which implies that good therapeutic response is less sensitive to perturbations and is therefore easier to achieve. Alternative NTCP models also exist (see *e.g.*, [54–57]) although most tend to be static (as opposed to dynamic, or temporal) and empirically founded.

Dynamic models of toxicity have also been developed. In [58] Krzyzanski et al. proposed a model of thrombocytopenia following combined chemotherapy and radiation treatment. Scenarios when the tumor model as well as the toxicity model are both dynamic can be approached using optimal control theory [59, 60]. The approach to selecting and ranking combinations presented in this paper could also be used in combination with other optimization approaches such as those that design treatment protocols to yield the most amount of information about the compounds [61, 62].

Conclusions and perspectives

We have demonstrated how a model-based approach, using TSE, can be used to compare and rank radiation and radiosensitizer combinations. The analysis weighs efficacy (tumor regression) against side-effects (toxicity) in order to provide a fair comparison and ranking of the different combinations.

While the chosen criteria for comparing combination therapies are natural, they are not the only reasonable choice. An alternative choice could be to compare the rate of tumor regression for each combination at a specified maximum tolerable exposure, *i.e.*, exchanging the roles that efficacy and toxicity play in the optimization problem.

Our analysis is focused on radiotherapy combined with radiosensitizing treatment. A similar approach using TSE and cost functions could also be considered for chemical combinations. However, the underlying pharmacokinetic,

pharmacodynamic, and toxicity modeling would have to account for potential drug interactions.

Funding Open access funding provided by Chalmers University of Technology. Tim Cardilin was supported by an education grant from Merck KGaA. This work was also partly funded by the Swedish Foundation for Strategic Research (Grant No. AM13-0046).

Declarations

Conflict of interest The authors have no conflicts of interest to declare that are relevant to the content of this article.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s10928-021-09784-7>.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Gong L et al (2021) Application of radiosensitizers in cancer radiotherapy. *Int J Nanomedicine* 16:1083–1102
- Liau SL et al (2013) New paradigms and future challenges in radiation oncology: an update of biological targets and technology. *Sci Transl Med* 5(173):173sr2
- Wang H et al (2018) Radiosensitizers. *Trends Pharmacol Sci* 39(1):24–48
- Hughes JP et al (2011) Principles of early drug discovery. *Br J Pharmacol* 162(6):1239–1249. <https://doi.org/10.1111/j.1476-5381.2010.01127.x>
- Swinney DC, Anthony J (2011) How were new medicines discovered? *Nat Rev Drug Discov* 10:507–519. <https://doi.org/10.1038/nrd3480>
- van der Greef J, McBurney RN (2005) Rescuing drug discovery: in vivo system pathology and systems pharmacology. *Nat Rev Drug Discov*. <https://doi.org/10.1038/nrd1904>
- De Ruyscher D et al (2019) Radiotherapy toxicity. *Nat Rev Dis Primers* 5(1):13
- Woodcock J et al (2017) Development of novel combination therapies. *N Engl J Med* 364:985–987
- Eking S et al (2007) In silico pharmacology for drug discovery: applications to targets and beyond. *Br J Pharmacol* 152(1):21–37
- Noori HR, Spanagel S (2013) In silico pharmacology: drug design and discovery's gate to the future. In *Silico Pharmacol*. <https://doi.org/10.1186/2193-9616-1-1>
- Sachs RK et al (2001) Simple ODE models of tumor growth and anti-angiogenic or radiation treatment. *Math Comput Model* 33:1297–1305. [https://doi.org/10.1016/S0895-7177\(00\)00316-2](https://doi.org/10.1016/S0895-7177(00)00316-2)
- Hong WS, Zhang GQ (2019) Simulation analysis for tumor radiotherapy based on three-component mathematical models. *J Appl Clin Med Phys* 20(3):22–26
- O'Rourke SFC et al (2009) Linear quadratic and tumour control probability modelling in external beam radiotherapy. *J Math Biol* 58(4–5):799–817
- Bodgi L et al (2016) Mathematical models of radiation action on living cells: from target theory to modern approaches. A historical and critical review. *J Theor Biol* 394:93–101. <https://doi.org/10.1016/j.jtbi.2016.01.018>
- Brenner DJ (2008) The linear-quadratic model is an appropriate methodology for determining iso-effective doses at large doses per fraction. *Semin Radiat Oncol* 18(4):234–239. <https://doi.org/10.1016/j.semradonc.2008.04.004>
- Kosinsky Y et al (2018) G. Radiation and PD-(L)1 treatment combinations: immune response and dose optimization via a predictive systems model. *J Immunother Cancer* 6(1):17
- Checkley S et al (2015) Bridging the gap between in vitro and in vivo: dose and schedule predictions for the ATR inhibitor AZD6738. *Sci Rep* 5:13545
- Begosh-Mayne D et al (2020) The dose–response characteristics of four NTCP models: using a novel CT-based radiomic method to quantify radiation-induced lung density changes. *Sci Rep* 10(1):10559
- Lin H et al (2012) Combining the LKB NTCP model with radiosensitivity parameters to characterize toxicity of radionuclides based on a multiclone kidney model: a theoretical assessment. *Australas Phys Eng Sci Med* 35(2):165–176
- Jakobi A et al (2015) Increase in tumor control and normal tissue complication probabilities in advanced head-and-neck cancer for dose-escalated intensity-modulated photon and proton therapy. *Front Oncol* 5:256
- Watanabe Y et al (2016) A mathematical model of tumor growth and its response to single irradiation. *Theor Biol Med Model*. <https://doi.org/10.1186/s12976-016-0032-7>
- Husband HR et al (2021) Model-based evaluation of image-guided fractionated whole-brain radiation therapy in pediatric diffuse intrinsic pontine glioma xenografts. *CPT Pharmacometrics Syst Pharmacol* 10(6):599–610
- Cardilin T et al (2018) Model-based evaluation of radiation and radiosensitizing agents in oncology. *CPT Pharmacometrics Syst Pharmacol*. <https://doi.org/10.1002/psp4.12268>
- Cardilin T et al (2019) Modeling long-term tumor growth and kill after combinations of radiation and radiosensitizing agents. *Cancer Chemother Pharmacol* 83(6):1159–1173
- Lyman JT (1985) Complication probability as assessed from dose-volume histograms. *Radiat Res Suppl* 8:S13–19
- Cardilin T et al (2017) Tumor static concentration curves in combination therapy. *AAPS J* 19(2):456–467. <https://doi.org/10.1208/s12248-016-9991-1>
- Jumbe NL et al (2010) Modeling the efficacy of trastuzumab-DM1, an antibody drug conjugate, in mice. *J Pharmacokinet Pharmacodyn* 37(3):221–242. <https://doi.org/10.1007/s10928-010-9156-2>
- Miao X et al (2016) Pharmacodynamic modeling of combined chemotherapeutic effects predicts synergistic activity of gemcitabine and trabectedin in pancreatic cancer cells. *Cancer Chemother Pharmacol* 77:181–193. <https://doi.org/10.1007/s00280-015-2907-4>
- Gabrielsson J et al (2016) Mixture dynamics: combination therapy in oncology. *Eur J Pharm Sci* 88:132–146. <https://doi.org/10.1016/j.ejps.2016.02.020>
- Adamus-Górka M et al (2011) Comparison of dose response models for predicting normal tissue complications from cancer radiotherapy: application in rat spinal cord. *Cancers (Basel)* 3(2):2421–2443

31. Källman P et al (1992) Tumour and normal tissue responses to fractionated non-uniform dose delivery. *Int J Radiat Biol* 62(2):249–262
32. Miller J et al (2009) The significance of the choice of radiobiological (NTCP) models in treatment plan objective functions. *Australas Phys Eng Sci Med* 32(2):81
33. Kutcher GJ, Burman C (1989) Calculation of complication probability factors for non-uniform normal tissue irradiation: the effective volume method. *Int J Radiat Oncol Biol Phys* 16(6):1623–1630
34. Keller F, Zellner D (1996) The 1-exp function as an alternative model of non-linear saturable kinetics. *Eur J Clin Chem Clin Biochem* 34:265–271
35. Almquist J et al (2015) Using sensitivity equations for computing gradients of the FOCE and FOCEI approximations to the population likelihood. *J Pharmacokinet Pharmacodyn* 42(3):191–209. <https://doi.org/10.1007/s10928-015-9409-1>
36. Williams MV et al (1985) A review of alpha/beta ratios for experimental tumors: implications for clinical studies of altered fractionation. *Int J Radiat Oncol Biol Phys* 11(1):87–96. [https://doi.org/10.1016/0360-3016\(85\)90366-9](https://doi.org/10.1016/0360-3016(85)90366-9)
37. Rationalizing Combination Therapies (2017) Editorial. *Nat Med* 23(10):1113. <https://doi.org/10.1038/nm.4426>
38. Webster RM (2016) Combination therapies in oncology. *Nat Rev Drug Discov* 15:81–82. <https://doi.org/10.1038/nrd.2016.3>
39. Jung J (2014) Human tumor xenograft models for preclinical assessment of anticancer drug development. *Toxicol Res* 30(1):1–5. <https://doi.org/10.5487/TR.2014.30.1.001>
40. EEPiA MID3 Workgroup et al (2016) Good practices in model-informed drug discovery and development: practice: application, and documentation. *CPT: Pharmacometrics Syst Pharmacol* 5(3):93–122. <https://doi.org/10.1002/psp4.12049>
41. Schuck E et al (2015) Preclinical pharmacokinetic/pharmacodynamic modeling and simulation in the pharmaceutical industry: an IQ consortium survey examining the current landscape. *AAPS J* 17(2):462–473. <https://doi.org/10.1208/s12248-014-9716-2>
42. Mould DR, Upton RN (2012) Basic concepts in population modeling, simulation, and model-based drug development. *CPT: Pharmacometrics Syst Pharmacol* 1:e6. <https://doi.org/10.1038/psp.2012.4>
43. Mould DR et al (2015) Developing exposure/response models for anticancer drug treatment: special considerations. *CPT Pharmacometrics Syst Pharmacol* 4(1):e00016. <https://doi.org/10.1002/psp4.16>
44. Mould DR, Hutson PR (2017) Critical considerations in anticancer drug development and dosing strategies: the past, present, and future. *J Clin Pharmacol* 57(Suppl 10):S116–S128. <https://doi.org/10.1002/jcph.983>
45. Simeoni M et al (2004) Predictive pharmacokinetic-pharmacodynamic modeling of tumor growth kinetics in xenograft models after administration of anticancer agents. *Cancer Res* 64(3):1094–1101
46. van Leeuwen CM et al (2018) The alpha and beta of tumours: a review of parameters of the linear-quadratic model, derived from clinical radiotherapy studies. *Radiat Oncol* 13(1):96
47. Loewe S (1953) The problem of synergism and antagonism of combined drugs. *Arzneimittelforschung* 3(6):285–290
48. Tallarida RJ (2001) Drug synergism: its detection and applications. *J Pharmacol Exp Ther* 298(3):865–872
49. Koch G et al (2016) Assessment of non-linear combination effect terms for drug-drug interactions. *J Pharmacokinet Pharmacodyn* 43(5):461–479. <https://doi.org/10.1007/s10928-016-9490-0>
50. Munro TR, Gilbert CW (1961) The relation between tumour lethal doses and the radiosensitivity of tumour cells. *Br J Radiol* 34:246–325
51. Bottino DC et al (2019) Dose optimization for anticancer drug combinations: maximizing therapeutic index via clinical exposure-toxicity/preclinical exposure-efficacy modeling. *Clin Cancer Res* 25(22):6633–6643
52. Baumann M, Petersen C (2005) TCP and NTCP: a basic introduction. *Rays* 30(2):99–104
53. Grégoire V (2005) Tumor control probability (TCP) and normal tissue complication probability (NTCP) in head and neck cancer. *Rays* 30(2):105–108
54. Lee TF et al (2012) Normal tissue complication probability model parameter estimation for xerostomia in head and neck cancer patients based on scintigraphy and quality of life assessments. *BMC Cancer* 12:567
55. Kinclová I et al (2020) Model-based calculation of thyroid gland normal tissue complication probability in head and neck cancer patients after radiation therapy. *Strahlenther Onkol* 196(6):561–568
56. Stieb S et al (2021) NTCP modeling of late effects for head and neck cancer: a systematic review. *Int J Part Ther* 8(1):95–107
57. Wals A et al (2006) Damage assessment in gastric cancer treatment with adjuvant radiochemotherapy: calculation of the NTCP's from the differential HDV of the organs at risk. *Clin Transl Oncol* 8(4):271–278
58. Krzyzanski W et al (2015) Pharmacodynamic model for chemoradiotherapy-induced thrombocytopenia in mice. *J Pharmacokinet Pharmacodyn* 42(6):709–720
59. Moore H (2016) How to mathematically optimize drug regimens using optimal control. *J Pharmacokinet Pharmacodyn* 45(1):127–137
60. Bruni C et al (2015) Optimal weekly scheduling in fractionated radiotherapy: effect of an upper bound on the dose fraction size. *J Math Biol* 71(2):361–398
61. Lestini G et al (2016) Optimal design for informative protocols in xenograft tumor growth inhibition experiments in mice. *AAPS J* 18(5):1233–1243
62. Floc'h N et al (2018) Optimizing the design of population-based patient-derived tumor xenograft studies to better predict clinical response. *Dis Model Mech* 11(11):dmm036160

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.